

Pfu/Psp DNA Polymerase 2X-preMix

Features:

Pfu/Psp DNA polymerase replicates DNA at 75°C catalyzing the polymerization of nucleotides into duplex DNA in the 5'=>3' direction in the presence of Mg⁺. Pfu DNA polymerase possesses 3' to 5' exonuclease proof reading activity that enables the polymerase to correct nucleotide-misincorporation errors. To reduce the risk of contamination, pipetting errors and to increase the repeatability of results the 2X-preMix contains an optimized mixture of enzyme, dNTP's and reaction buffer. Just add your template DNA and primers.

Applications:

- blunt end PCR cloning
- PCR and primer extension where "high fidelity" is required
- Site-directed mutagenesis
- PCR where visual control is needed

Description:

Pfu/Psp DNA polymerase 2X-preMix is isolated from the archae bacteria *Pyrococcus f*-species, a thermostable Polymerase of approximately 90000 daltons. Base misinsertions that may occur during polymerization are rapidly excised by the proofreading activity of the polymerase. The Pfu/Psp DNA Polymerase has **no detectable reverse transcriptase activity**.

Concentration: Premix 2X (25µl per reaction); the final concentration of Mg⁺ is 2 mM.

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Unit definition:

One unit is defined as the amount of enzyme required to catalyze the incorporation of 10 nM of dNTPs into acid insoluble material in 30 minutes at 75°C.

Quality control:

- Tested for the DNA amplification of 2,2 kb from lambda DNA
- Contamination level check of bacterial DNA
- Purity by SDS-Page > 90 %

Usage:

Standard protocol:

- Do not use dUTP or dITP or primers containing these nucleotides

| Components | Volume per reaction | end conc. |
|---------------------------------------|---------------------|------------|
| Up-stream primer (e.g. 20 µM) | 0,5 µl | 0.1-1.0 µM |
| Down-stream primer (e.g. 20 µM) | 0.5 µl | 0.1-1.0 µM |
| Template DNA (10 ng/µl) | 1.0 µl | <= 0,5 µg |
| Pfu/Psp 2X-preMix | 25 µl | 1X |
| Sterile dest. Water (molecular grade) | up to 50 µl | |

Note:

- vortex all solutions carefully before using
- dispense all reagents on ice to avoid degradation of primers and dNTP's
- add the enzyme after Template DNA
- may you have to optimize the Mg⁺ concentration for best result

General Thermo-Cycler protocol:

| Step | Time | Temperature |
|----------------------|-------------------|-------------|
| Initial denaturation | 1-3 min | 95°C |
| 25-35 Cycles: | | |
| Denaturation | 30-100 sec | 95°C |
| Annealing | 30-65 sec | 37-69°C |
| Extension | 1-2 min (per 1kb) | 72-75°C |
| Final extension | 5 min | 72-75°C |

Loading on the gel:

Recommended volume is 10 µl of reaction mixture

Storage: at -20°C for 24 months

Transportation: on blue ice

Ordering Information:

| Cat.-no | Description | Amount |
|---------|----------------------------------|-------------|
| S121 | Pfu/Psp 2X-preMix DNA Polymerase | 2 x 50 rcs |
| S122 | Pfu/Psp 2X-preMix DNA Polymerase | 10 x 50 rcs |

.. a good decision ..